# Modeling proteins based on homology

Exercise 1: Homology detection & structure prediction of an uncharacterized protein from *Modestobacter marinus* using HHpred

1. Retrieve the query sequence:



You will work with a uncharacterized protein from *Modestobacter marinus*. Download the protein sequence of in fasta format and save into a directory. Open **http://www.uniprot.org/** in your browser, and search for the gene name *MODMU\_2448*. (Uniprot Id : *I4EWW4*)

2. Find template structures:

Standard Protein BLAST

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We can template structures by performing a BLAST search against the Protein Data Bank 'PDB'.

Open the Ncbi Blast page in your browser. https://blast.ncbi.nlm.nih.gov/Blast.cgi

Paste your FASTA sequence into the box Enter a title and use default parameters Choose 'pdb' as your search database Run BLAST

Did you find any templates?. [Hint: There are no templates available in PDB for this protein].

#### 3. Find template structures by fold recognition:

Now we can try to use 'fold recognition' methods to find potential template structures. We are going to use the **'HHpred'** server to search for protein folds. HHpred was used for sequence and structure database searching to find remote homologs.

Input							
Paste protein sequence or MSA							
or upload a local file Select input format	Search with pancreatitis ind Browse No file sele FASTA						
			Reset form	Submit job			
Search Options							
Select HMM databases (hold Ctr to select several)	Standard pdb70_04Mar17 pdb_on_hold_7Mar17 mmcif70_Dec16 SCOPe95_2.06 SCOPe70_2.06	Proteomes Arabidopsis_thaliana Caenorhabditis_elegans Drosophila_melanogaster Homo_sapiens Mus_musculus					
MSA Generation Method	🗿 HHblits 🔵 Psiblast						
Max. MSA Generation iterations	3 -						
Score secondary structure	O yes ○ no ○ predicted vs predicted only						
Alignment mode	💿 local 🔵 global						
Realign with MAC							

HHpred - Homology detection & structure prediction by HMM-HMM comparison

Open the HHpred web server. https://toolkit.tuebingen.mpg.de/hhpred

Past your target sequence in fasta format and choose default parameters for other options. Choose *pbd70* for the option '**Select HMM Databases**'. Submit Job and wait for the results.

### 4. HHpred - Results

Analyze the results. The output is organized into three sections:

1. A bar graph summarizing the position

2. A tabular hit list with probabilities, E-values, scores, and match regions in query and templates.

3. The pairwise query-templates alignments

It is not possible to identify these templates by just sequence comparison. Note the poor percent identity of the alignment for the top hits.

#### 5. Build model

From the HHpred results page click Results ---> Create model.

HHpred - Results Job-ID: 2441985 Date: 07:48 on Mar 08 2017									
Results	Histogram	Reduced alignment	Representative alignment	Full alignment					
Create m	odel Merge	Q/T alignments For	ward to PCoils Forward MSA	Save Export					
1. M th M	anual tem le selected ODELLER.	plate selection: Ge d template or te	enerate a PIR-alignmen mplates in order to t	t of your sequ puild a 3D n	uence with nodel with	Create model from manually selected template(s)			
2. Au Hi m	utomatic t MMs, rerar ultiple alig	emplate selection Ik templates and a nment is created f	: Optimize diversities automatically select bes rom this set, and a 3D n	of query and t set. In furth nodel is build v	l template her steps a with MODEL	Automatically select best templates			
	Only hits fo	ound in PDB or SCC	OP can be used to create	a model (oth	er hits are d	lisabled).			

. . . .

HHpred will connect to modeller and do the model building for you!. You can manually or ask hhpred to automatically select template from the results.

To select template manually, scroll down to tabular hit list and check the check box for the template of your preference. Then click 'create model from manually selected template(s)'. This will open the modeller web server page with the alignment already pasted in the box. You need to just submit the modeller job.

Enter your Modeller-key Choose a Job-id Input notification email Submit Job.

#### 6.Results

The result page displays the modelled structure pdb-file. You can choose pdb-file  $\rightarrow$  save to save the model in pdb format.

Now use chimera to view your model and compare it with the template structure.

[Answer: The uncharacterized protein contains the ADENYLATE CYCLASE fold]

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Exercise 2: Homology detection & structure prediction of an uncharacterized protein from *Candidatus Entotheonella* using HHpred

You will work with a uncharacterized protein from *Candidatus Entotheonella*. Download the protein sequence of in fasta format and save into a directory. Open **http://www.uniprot.org/** in your browser, and search for the gene name *ETSY2\_32710*. (Uniprot Id : W4M0R9)

Repeat the same steps as Exercise 1.

Hint: Look at the template length and coverage for HHpred hits. You may need to choose more than one template for modelling.

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Exercise 3: Homology detection & structure prediction of an uncharacterized protein of your interest

## 1. Retrieve the query sequence:



You will work with a uncharacterized protein of your interest.

Open http://www.uniprot.org/ in your browser, and choose 'Advanced' in the search bar.

	Searching in UniProtKB	Help			×
		Protein name [DE]	Term Incharacterized protein		
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Enter 'Uncharacterized protein' for protein name and search. One you get the results you can choose any protein of your interest. Note that you can sort the results on the results page by sequence length, organism name etc.

Hint: You have more chance of finding a template for proteins with sequence length < 400, Also it is faster to build the model for smaller proteins.

Once you select your protein, repeat the same steps as Exercise 2.

Modeling proteins using automated modelling servers

Exercise 4: Automated protein structure prediction using Phyre2

## Go to Phyre2 web server

http://www.sbg.bio.ic.ac.uk/phyre2/html/page.cgi?id=index



Phyre2 is fully automated, you just need to give your protein sequence for modelling. The program will find the template, create alignment and build the model.

Use any of the previous example to build model and compare the results.

Exercise 5: Automated protein structure prediction using I-TASSER

Go to I-TASSER web server http://zhanglab.ccmb.med.umich.edu/I-TASSER/